

Synthesis of an 11-Unsubstituted Analogue of (±)-Huperzine A

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Abstract

The synthesis of a new analogue of (±)-huperzine A lacking the ethylidene appendage at position 11 has been accomplished. The key steps of the synthesis were: a) the reduction of the keto function of the known methyl 7.7-ethylenedioxy-3-methyl-9-oxobicyclo[3.3.1]non-3-ene-1-carboxylate into a methylene group by reduction to a mixture of stereoisomeric alcohols followed by alcohol-deoxygenation through a Barton-McCombie procedure and b) the elaboration of the pyridone ring in a late stage of the synthesis by reaction of a pyrrolidine enamine with propiolamide, which gave a mixture of regioisomeric pyridone derivatives. © 1999 Elsevier Science Ltd. All rights reserved.

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Alzheimer's disease (AD) is the most common cause of dementia in the aged [1]. Most neurotransmitter systems have been implicated in the etiology of AD, the cholinergic neurotransmission being specially affected [2-6]. Accordingly, enhancement of the central cholinergic function, for example by means of reversible acetylcholinesterase (AChE) inhibitors [7,8], is one of the most promising methods for the symptomatic treatment of AD.

Huperzine A (1) a lycopodium alkaloid, isolated from the club moss *Huperzia serrata* (Thunb.) Trev. = *Lycopodium serratum* Thunb., a Chinese traditional medicine [9-11], is a potent and selective reversible inhibitor of AChE which appears to be superior to other AChE inhibitors such as tacrine, physostigmine or galanthamine, because of its comparatively longer duration of action and higher therapeutic index [12]. Huperzine A has been considered an important lead compound in the search for better cholinomimetics for the treatment of AD. The low natural abundance of huperzine A induced several groups to develop synthetic routes to this compound [13-19] and to prepare huperzine A analogues with the aim of studying the effects of structural modifications on the biological activity (structure-activity relationships).

Figure 1. Huperzine A (1), tacrine (2), and related compounds.

In this sense, numerous huperzine A analogues have been prepared by different modifications of the parent structure. Structural modifications investigated to date include those carried out at the pyridone ring (replacement by an unsubstituted benzene ring [20], a monohydroxy and a dihydroxy-substituted benzene ring [21] or different heterocyclic rings [22,23], opening of the pyridone ring and replacement of the amide function with an isosteric carbamate group [24], alkylation of the nitrogen atom [25]), at the amino group (alkylation [16], replacement by an aminomethyl group [16]), at the ethylidene group (isomerization [26], replacement by a methylene [16], propylidene [16] or by other alkylidene or alkyl groups [17, 27-29]), at the C-12 methyl group (replacement by a hydrogen atom [30] or by a phenyl [16], halomethyl [28,29], hydroxymethyl [28] or alkoxycarbonyl [28] group), at the unsaturated three-carbon bridge (saturation [31] or isomerization of the endocyclic double bond to an exocyclic position [32], cyclopropanation [33], hydration [25]); at the C-10 position (introduction of axial and / or equatorial alkyl groups [34-36]) as well as those carried out by extensive simplification of the carbobicyclic moiety [30,31,37]. It is worth noting that most of these analogues were found not to exhibit useful AChE inhibitory activity. To the best of our knowledge, the sole analogues which can rival the activity of the parent compound are the C-10 axial methylated, the 10,10dimethylated, and the 10-spirocyclopropyl derivatives.

Recently, we published the synthesis and evaluation of a series of compounds, designed by combining the pharmacophores of huperzine A (1) (carbobicyclic substructure) and tacrine (THA) (2) (4-aminoquinoline substructure) [38]. Among these THA-huperzine A hybrids, compound 3, the sole compound of the series which incorporated the ethylidene group of huperzine A, turned out to be 2.5-fold less potent than THA as an AChE inhibitor, while compound 4, lacking the ethylidene substituent, was 2-fold more potent than THA. From the chemical and biological data reported to date it would appear that the ethylidene group is one of the essential features for high AChE inhibitory activity in huperzine A analogues (see above). However, the biological data obtained from our THA-huperzine A hybrids seem to indicate that, in this kind of compound, the presence of this ethylidene group is not necessary for a better AChE inhibitory activity. In order to ascertain the relevance of the ethylidene substituent at position 11 to the biological activity of huperzine A analogues, we embarked upon the synthesis and evaluation of a new analogue of (±)-huperzine A lacking this substituent (5).

Results and Discussion

We describe herein the synthesis of the new (±)-huperzine A analogue 5 from the known keto ester 7, which was prepared in 34% overall yield from commercially available

i) Ni₂B, NaBH₄, MeOH, r.t., 30 min, 24% yield of 8 and 26% yield of 9; ii) LiBHEt₃, THF, r.t., 30 min, 58% yield of 8 and 29% yield of 9 based on recovered 7; iii) 80% aq. NH₂NH₂·H₂O, KOH, diethylene glycol, 140 °C, 17 h; iv) Conc. HCl, r.t., 5 min. 40% overall yield of 11 from 7; v) NaBH₄, MeOH, r.t., 24 h, 31% yield; vi) 1,1'-Thiocarbonyldiimidazole, 1,2-dichloroethane, reflux, 3 h, 71% yield of 13; vii) Bu₃SnH, AIBN, toluene, reflux, 1.5 h, 42% yield of 10 from 13 and 60% overall yield of 10 from 9 via 14.

Scheme 1. Reduction of keto ester 7 to 10.

1,4-cyclohexanedione monoethylene ketal 6, following an improved procedure developed by our group [39]. The main features of the synthetic plan are: 1) the deoxygenation of the keto function of 7 to give 10, 2) the conversion of the ester function of 10 into a protected amino group, 3) the elaboration of the pyridone ring, and 4) the deprotection of the amino function.

The reduction of keto ester 7 to 10 proved to be a rather difficult task (Scheme 1). For this transformation, only alkaline or neutral conditions could be used, due to the presence of acidsensitive functionalities, i.e. C=C double bond and ketal group. Treatment of 7 under standard Wolff-Kishner reduction conditions [40] did not afford the expected methylenic product. pyrazolinone 11 being formed instead in about 40% yield. The formation of 11 might be easily explained by intramolecular nucleophilic addition of the amino group of an initially formed hydrazone to the ester function, followed by ketal hydrolysis during the acid work-up. Thus, we decided to prepare ester 10 through a two-step sequence which involved initial reduction of the keto group to an alcohol followed by removal of the hydroxy group by a Barton-McCombie [41] deoxygenation reaction. Reduction of keto ester 7 with an excess of NaBH₄ in MeOH at room temperature gave diol 12 in 31% yield, as the sole isolated product. The same reaction carried out by using a limited amount of NaBH₄ gave a complex mixture of products. Attempted reduction of 7 with an equimolar amount of NaBH₃CN left the starting compound unchanged. Finally, we were able to selectively reduce the keto group of keto ester 7 with nickel boride / NaBH₄ [42-44]. Reaction of keto ester 7 with NiCl₂·6H₂O and NaBH₄, in the molar ratio of 1:1.1:2.5, afforded a mixture of diastereomeric alcohols 8 and 9, which was separated by column chromatography (24% and 26% yield, respectively). Moreover, reaction of 7 with lithium triethylborohydride (Super-Hydride®) [45] in THF at room temperature for 30 min afforded hydroxy esters 8 and 9 in better overall yield (50% and 25% yield, respectively; 58% and 29%, respectively, based on recovered starting material).

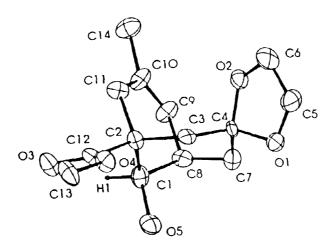


Figure 2. Crystal structure (ORTEP) of hydroxy ester 9.

The configuration of hydroxy esters 8 and 9 was established through the analysis of their ¹³C NMR spectra (Table 1). As can be seen from Table 1, the signals of the C6 and C8 carbon atoms of the *syn*-hydroxy ester 9, in which the hydroxy group is *axial* with respect to the dioxolane-bearing cyclohexane, appear at 6.2 and 5.5 ppm, respectively, shifted upfield compared with the corresponding signals of the *anti*-derivative 8, due to the γ-gauche effect of the hydroxy group. For the same reason, the signals of the C2 and C4 carbon atoms of hydroxy ester 8 appear at 5.7 and 4.0 ppm, respectively, shifted upfield as compared with the corresponding signals of hydroxy ester 9. Comparison of the chemical shifts of C2, C4, C6 and C8 of diol 12 with the corresponding values of hydroxy esters 8 and 9, show very small differences due to the change of the methoxycarbonyl for the hydroxymethyl group (0.1-0.3 ppm), when compared with hydroxy ester 8 and much larger differences (4.1-6.0 ppm) when compared with 9, in accord with an *anti* arrangement of the secondary hydroxy group of diol 12. X-ray diffraction analysis of hydroxy ester 9 (Figure 2) allowed us to clearly establish the *syn* arrangement of its hydroxyl group. In Table 3 further crystallographic data of this compound are compiled.

The removal of the hydroxy group of hydroxy esters 8 and 9 was carried out through a Barton-McCombie deoxygenation procedure [41] (Scheme 1). Hydroxy ester 8 was easily converted into the corresponding thiocarbonyl imidazolide 13 in 71% yield, on reaction with 1,1'-thiocarbonyldiimidazole in 1,2-dichloroethane under reflux. However, the conversion of 9 into the imidazolide 14 was not so efficient. Treatment of the more hindered hydroxy ester 9 with 1,1'-thiocarbonyldiimidazole under the same reaction conditions gave imidazolide 14 in only 12% yield, much of the starting compound being recovered unchanged (70%). When the above reaction was carried out in a pressure flask at a higher temperature (120 °C), unreacted 9 was recovered in 50% yield while imidazolide 14 was obtained in somewhat better yield (32%; 64% yield based on recovered 9). Compound 14 was not stable enough to allow its complete characterization and was used as such in the next step.

The second step in the deoxygenation procedure implied the radical chain reduction of imidazolides 13 and 14 with Bu₃SnH [41]. In terms of initiator requirement, it has been reported that initiators are not usually necessary for substrates such as 13 and 14, possessing thiocarbonyl imidazolide functionality in secondary alcohols [46]. In practice, reactions of imidazolides 13 and 14 with Bu₃SnH in refluxing toluene furnished ester 10 in only moderate to low yields (13% and

i) 20% aq. NaOH, THF / MeOH 1:1, reflux, 48 h; ii) 5 N HCl, r.t., 5 min, 54% overall yield of 15 from 10; iii) (C₆H₅O)₂P(O)N₃. Et₃N, chlorobenzene, 90 °C, 3.5 h; iv) MeOH, reflux, 17 h, 99% overall yield of 16 from 15; v) Pyrrolidine, 4 Å molecular sieves, benzene, reflux, 5 h; vi) Propiolamide, reflux, 15 h, 28% overall yield of 19 and 27% overall yield of 20 based on recovered 16; vii) MPLC separation; viii) *n*-PrSLi, HMPA, 40 °C, 24 h, 81% yield based on recovered 19; ix) (CH₃)₃SiI, CHCl₃, reflux, 8 h; x) McOH, reflux, 14 h, 66% overall yield of 21 from 20.

Scheme 2. Synthesis of the huperzine A analogues 5 and 21.

45% yield, respectively). However, these deoxygenation reactions proceeded more efficiently on addition of AIBN (0.1 equiv.) as initiator of the radical chain reduction. In this way, imidazolides 13 and 14 were deoxygenated to ester 10 in 42% and 60% yield, respectively.

Next, we undertook the introduction of a precursor of the bridgehead amino group through a Curtius rearrangement. To this end, ketal ester 10 was first converted into keto acid 15, by saponification of the hindered ester group with 20% aqueous NaOH in a mixture of THF and MeOH in the ratio of 1:1 under reflux for 48 h [47], followed by hydrolysis of the ketal function during the acid work-up (Scheme 2). Curtius rearrangement of keto acid 15 was carried out by reaction with diphenylphosphoryl azide [48] followed by methanolysis of the resulting isocyanate, keto carbamate 16 being obtained in almost quantitative yield.

Elaboration of the pyridone ring was carried out by using a modification of the procedure described by Kozikowski *et al.* [49]. Keto carbamate 16 was reacted with pyrrolidine in refluxing benzene in the presence of 4 Å molecular sieves and the product thus obtained was heated under reflux with propiolamide, previously prepared from ethyl propiolate and NH₄OH [50]. This reaction provided an approximately 1:1 mixture of the regioisomeric pyridones 19 and 20, which were partially separated by flash column chromatography, a significant amount of starting 16 (40%) being recovered (55% total yield of pyridones 19 and 20, based on recovered 16) (Scheme 2). These pyridones were efficiently separated by medium pressure liquid chromatography (MPLC) through silica gel (40-60 μm) using mixtures of AcOEt / MeOH as eluent. Although the *anti*-enamine 17 must be formed preferentially, as was observed for a related case [24], a significant amount of the *syn*-pyridone 20 was obtained in this reaction. Calkylation of *anti*-enamine 17 with propiolamide should be slower than C-alkylation of the *syn*-enamine 18 due to the steric effect of the substitutent at the bridgehead position. Isomerization of enamine 17 to 18 under the reaction conditions could explain the formation of the regioisomeric pyridone 20.

Cleavage of the methyl carbamate of 19 was accomplished without problems by reaction with lithium 1-propanethiolate in hexamethylphosphoramide [48], obtaining the huperzine A analogue 5 in 63% yield (81% yield, based on recovered 19). However, cleavage of the carbamate group of 20 under the same reaction conditions led to a complex mixture of products not containing the expected amine. Also, attempted hydrolysis of the carbamate group of 20 by reaction with potassium hydroxide in the presence of 18-crown-6 [51] failed. Finally, treatment of 20 with trimethylsilyl iodide (TMSI) followed by reaction with methanol under reflux [47] gave mainly 21, a product in which not only the carbamate function had been hydrolyzed, but also the C=C double bond had isomerized to a neighbouring position. This isomerization can be easily explained under acid catalysis (some HI formed by hydrolysis of the TMSI) and reflects the greater stability of the *anti*-arrangement for the C=C double bond and the heterocyclic ring in this kind of bicyclo[3.3.1]nonadiene derivative. A similar situation was observed [24] in related cases.

Except for pyrazolone 11 and imidazolide 14, the new compounds herein described have been fully characterized though their spectroscopic data and elemental analysis or high resolution mass spectrometry (5 and 21), and by X-ray diffraction analysis in the case of hydroxy ester 9.

Assignment of the ¹H and ¹³C NMR spectra was carried out in a standard way with the aid of DEPT, ¹H / ¹H and ¹H / ¹³C COSY experiments.

The methodology herein described for the synthesis of compound 5 is currently being applied to the synthesis of other huperzine A analogues, whose biological activity will be reported in due course.

Experimental

General. Melting points were determined with a MFB 595010 M Gallenkamp melting point apparatus. 500 MHz ¹H NMR spectra were performed on a Varian VXR 500 spectrometer, 75.4 MHz ¹³C NMR spectra on a Varian Gemini 300, and 200 MHz ¹H and 50.3 MHz ¹³C NMR spectra on a Varian Gemini 200. Except where otherwise stated, ¹H NMR spectra were recorded at 500 MHz and ¹³C NMR spectra at 75.4 MHz, in CDCl₃. Chemical shifts (δ) are reported in ppm related to internal tetramethylsilane. Assignments given for the NMR spectra are based on DEPT, ¹H/¹H and ¹H/¹³C COSY experiments (HMQC sequence). The ¹³C and ¹H NMR data of compounds 8-10, 12, 13, 15, and 16 are collected in Tables 1 and 2, respectively. while those of compounds 5, 11, and 19-21 are described in the experimental. IR spectra were recorded on a FT/IR Perkin-Elmer spectrometer, model 1600. Routine MS spectra were taken on a Hewlett-Packard 5988A spectrometer, by direct introduction of the sample, using the electron impact technique (70 eV). Significant ions given are those showing the following relative abundances: 5% or more if m/z > 150 and 25% or more if $m/z \le 150$. Silica gel SDS 60 (60-200) µm) was usually utilized for the column chromatography. Medium pressure liquid chromatography (MPLC) separations were carried out on a MPLC equipment which consisted of a pump (Büchi 688) a variable λ detector (Büchi) and a column (45 \times 3.5 cm) containing silica gel (40-60 µm) as the stationary phase. Elemental analyses and high resolution mass spectra were carried out, respectively, at the Microanalysis Service and the Mass Spectrometry Laboratory of the Centro de Investigación y Desarrollo (C.I.D.), C.S.I.C., Barcelona, Spain.

Methyl 7,7-ethylenedioxy-anti-9-hydroxy-3-methylbicyclo[3.3.1]non-3-ene-1-carboxylate (8) and methyl 7,7-ethylenedioxy-syn-9-hydroxy-3-methylbicyclo[3.3.1]non-3-ene-1-carboxylate (9).

By reduction with Ni₂B / NaBH₄. To a stirred suspension of NiCl₂·6H₂O (977 mg, 4.11 mmol) in MeOH (80 mL), NaBH₄ (106.2 mg, 2.79 mmol) was added portionwise over a 5-minute-period. The resulting suspension was stirred thoroughly for 2 h and a solution of keto ester 7 (1.00 g, 3.76 mmol) and NaBH₄ (250 mg, 6.58 mmol) in MeOH (20 mL) was added dropwise over 5 min. The reation mixture was stirred for 30 min and filtered through Celite[®], washing the solid with MeOH (50 mL). The combined filtrate and washings were treated with 25% aqueous NH₄OH (100 mL), diluted with water (100 mL), and extracted with CH₂Cl₂ (4 × 100 mL). The combined organic extracts were washed with brine (100 mL), dried with anhydrous Na₂SO₄ and concentrated *in vacuo* to give a colorless oily residue (670 mg), which was submitted to column chromatography through silica gel (67 g, hexane / AcOEt, gradient elution). On elution with hexane / AcOEt 1:1, alcohol 9 (190 mg), a mixture of alcohols 9 / 8 (240 mg) in a ratio 7:16 (¹H NMR), and alcohol 8 (70 mg) were successively separated (24% total yield of 8 and 26% total yield of 9). The analytical samples of 8 and 9 were obtained by distillation at 130 °C / 1 Torr and 150 °C / 2 Torr, respectively.

Spectroscopic and analytical data of alcohol 8: colorless oil. IR (NaCl) v 3493, 2952, 2916, 1732, 1434, 1356, 1254, 1231, 1151, 1140, 1094, 1054, 998, 979, 950, 826 cm⁻¹. MS, m/z (%): 269 (8), 268 (M·+, 33), 251 (7), 250 (M·+ - H₂O, 29), 237 (5), 236 (M·+ - CH₃OH, 26), 218 (6), 210 (5), 209 (M+ - COOMe, 26), 208 (8), 207 (5), 206 (7), 192 (12), 191 (M+ - COOMe - H₂O, 70), 188 (8), 179 (6), 178 (5), 175 (18), 174 (45), 167 (23), 166 (M·+ - C₅H₁₀O₂, 76), 165 (22). 164 (19), 163 (10), 157 (9), 154 (13), 153 (10), 152 (14), 151 (9), 147 (M+ - COOMe - H₂O - C₂H₄O, 49), 135 (40), 134 (66), 119 (53), 107 (34), 105 (74), 95 (37), 93 (37), 91 (82), 87 (96). 86 (94), 81 (32), 80 (48), 79 (79), 77 (81), 67 (46), 65 (43), 59 (100), 55 (89), 53 (59), 51 (25). Anal. Calcd. for C₁4H₂O₅: C, 62.67; H, 7.52. Found: C, 62.74; H, 7.72.

Spectroscopic and analytical data of alcohol **9**: white crystals, m.p. 79-81 °C. IR (KBr) v 3462, 3044, 2960, 2919, 2890, 1696, 1445, 1427, 1297, 1273, 1240, 1188, 1143, 1113, 1094, 1070, 1005, 987, 956, 938, 832, 802 cm⁻¹. MS, m/z (%): 268 (M·+, 15), 250 (M·+ - H₂O, 10), 209 (M+ - COOMe, 25), 192 (6), 191 (M+ - COOMe - H₂O, 32), 188 (10), 175 (9), 174 (15), 166 (5), 165 (M+ - COOMe - C₂H₄O, 14), 164 (13), 163 (6), 157 (11), 154 (11), 153 (5), 152 (17), 151 (9), 147 (M+ - COOMe - H₂O - C₂H₄O, 36), 121 (27), 119 (49), 107 (25), 105 (46), 95 (29), 93 (33), 91 (71), 87 (78), 86 (74), 81 (31), 80 (56), 79 (75), 77 (71), 73 (25), 67 (48), 65 (41), 59 (67), 57 (32), 55 (100), 53 (62), 51 (25). Anal. Calcd. for C₁4H₂0O₅: C, 62.67; H, 7.52. Found: C, 62.61; H, 7.62.

By reduction with lithium triethylborohydride. A solution of keto ester 7 (3.50 g, 13.2 mmol) in anhydrous THF (220 mL) was treated dropwise with lithium triethylborohydride (1 M solution in THF, 19.8 mL, 19.8 mmol) over 5 min. The reaction mixture was stirred at room temperature for 30 min, treated with saturated aqueous NH₄Cl (250 mL), diluted with water (350 mL), and extracted with CH₂Cl₂ (5 × 150 mL). The combined organic extracts were dried with anhydrous Na₂SO₄ and concentrated *in vacuo* to give a yellowish oil (4.80 g), which was

Table 1
13C NAP Chamical Shifte al of Compounds 8-10 12 13 15 and 16

| C NMR Chemical Shifts ¹⁴ of Compounds 8-10, 12, 13, 15, and 16. | | | | | | | |
|--|------------|------------|------------|------------|-------------------|-------|-------|
| | 8 | 9 | 10 | 12[b,c] | 13 ^[d] | 15 | 16 |
| CI | 46.9 | 47.0 | 42.4 | 38.5 | 45.9 | 44.6 | 54.1 |
| C2 | 34.4 | 40.1 | 38.7 | 34.1 | 35.5 | 39.3 | 44.0 |
| C3 | 134.7 | 132.9 | 133.5 | 136.8 | 134.1 | 131.9 | 131.9 |
| C4 | 118.5 | 122.5 | 123.5 | 118.4 | 118.4 | 124.0 | 124.4 |
| C5 | 36.5 | 35.3 | 29.6 | 38.0 | 32.9 | 31.0 | 31.0 |
| C6 | 38.7 | 32.5 | 38.6 | 38.4 | 38.1 | 45.6 | 45.2 |
| C7 | 107.5 | 108.2 | 108.7 | 108.2 | 106.8 | 209.6 | 209.2 |
| C8 | 42.9 | 37.4 | 43.4 | 42.6 | 43.9 | 50.0 | 52.9 |
| C9 | 70.8 | 68.9 | 33.0 | 73.8 | 81.7 | 32.4 | 35.0 |
| 3-CH ₃ | 22.8 | 22.4 | 22.9 | 23.0 | 22.7 | 22.9 | 22.6 |
| OCH ₃ | 52.1 | 52.1 | 52.0 | | 52.5 | | 51.7 |
| OCH ₂ CH ₂ O | 62.8, 64.8 | 62.9, 64.8 | 62.8, 64.9 | 62.8, 64.8 | 63.1, 65.0 | | |
| coo | 176.5 | 177.6 | 177.8 | | 174.2 | 182.0 | 155.1 |

[[]a] All these spectra were taken at 75.4 MHz in CDCl3.

¹H NMR Chemical Shifts^[a,b] and Coupling Constants of Compounds 8-10, 12, 13, 15, and 16.

| | 8 | 9 | 10 | 12[c,d] | 13[e] | 15 ^[f] | 16 g |
|------------------------------------|-----------|-----------|-----------|-----------|-----------|-------------------|-----------|
| 2-Hexo | 2.09 | 2.24* | 2.34 | 2.33 | 2.65 | 2.59 | 2.20* |
| 2-Hendo | 2.52 | 2.29* | 2.05 | 1.75 | 2.32 | 2.02 | 2.28* |
| 4-H | 5.27 | 5.35 | 5.44 | 5.25 | 5.24 | 5.43 | 5.38 |
| 5-H | 2.54 | 2.46 | 2.50 | 2.48 | 2.98 | 2.77 | 2.74 |
| 6-Hexo | 1.91 | 2.16 | 1.67-1.77 | 1.84 | 2.05 | 2.42 | 2.43 |
| 6-H <i>endo</i> | 1.81 | 1.54 | 1.67-1.77 | 1.80 | 1.87 | 2.33 | 2.27 |
| 8-Hexo | 1.92 | 2.19 | 1.84 | 1.52 | 2.13 | 2.69 | 3.00 |
| 8-Hendo | 1.92 | 1.81 | 1.97 | 1.48 | 2.01 | 2.46 | 2.53 |
| 9-Hsyn | 4.15 | | 1.67-1.77 | 3.78 | 5.99 | 2.19 | 2.44-2.50 |
| 9-Hanti | | 4.15 | 1.85 | | | 2.13 | 2.02 |
| 3-CH ₃ | 1.70 | 1.61 | 1.63 | 1.72 | 1.74 | 1.62 | 1.59 |
| OCH ₃ | 3.71 | 3.70 | 3.66 | | 3.65 | | 3.62 |
| OCH ₂ CH ₂ O | 3.73-3.97 | 3.73-3.98 | 3.70-3.97 | 3.69-3.95 | 3.72-3.98 | | |
| 9-OH | 2.60 | 3.10 | | 1.90-2.30 | | | |
| J (Hz) | | | | | | | |
| 2-Hexo/2-Hendo | 18.5 | 17.5 | 18.0 | 18.5 | 18.5 | 17.5 | 17.0 |
| 2-Hexo/8-Hexo | | | 1.5 | | 1.0 | 1.5 | |
| 2-Hendo/9-Hsyn | | | | | 1.0 | | |
| 4-H/5-H | 6.0 | 6.5 | 6.5 | 6.0 | 5.5 | 4.5 | 4.5 |
| 5-H/6-Hexo | 4.5 | 5.0 | | 4.5 | 4.5 | 4.0 | 4.5 |
| 5-H/6-Hendo | 2.0 | 2.5 | | | 3.0 | 3.0 | 2.5 |
| 5-H/9-Hsyn | 3.0 | | | | 3.5 | | |
| 5-H/9-Hanti | | | 3.5 | | | 4.0 | 4.0 |
| 6-Hexol6-Hendo | 14.0 | 14.0 | | 14.0 | 14.0 | 14.5 | 14.5 |
| 6-Hendo/8-Hendo | 2.0 | 2.5 | 2.0 | 2.0 | 3.0 | 2.0 | 2.0 |
| 6-Hendol9-Hanti | | | 1.5 | | | 2.0 | 2.0 |
| 8-Hexo/8-Hendo | | 14.0 | 13.5 | 14.0 | 14.0 | 15.5 | 15.5 |
| 8-Hendo/9-Hanti | | 1.0 | 2.0 | | | 2.0 | 2.0 |
| 9-Hsyn/9-Hanti | | | 12.5 | | | 13.0 | 12.5 |
| 9-H /9-O H | 6.5 | 2.0 | | 3.0 | | | |

[[]a] All these spectra were taken at 500 MHz in CDCl3.

[[]b] 12 has been named as a methanol derivative to keep the same numbering as the rest of compounds of this Table.

[[]c] Another signal: CH2OH 70.8.

[[]d] Other signals: C=S 182.8, C2' 136.4, C4' 130.6, and C5' 117.8.

[[]b] The values indicated with * within a column can be interchanged.

[[]c] Other signals: CH2OH 3.38 and 3.49, CH2OH 2.50-3.10.

[[]d] Another coupling constant: CH2OH $J_{gem} = 11.0 \text{ Hz}$.

[[]e] Other signals: 2'-H 8.13, 4'-H 6.97, 5'-H 7.46.

[[]f] Another signal: COOH 9.0-12.0.

[[]g] Another signal: NH 4.75.

taken up in CH₂Cl₂ (250 mL) and washed with water (125 mL). The organic layer was dried with anhydrous Na₂SO₄ and evaporated at reduced pressure, to give a yellowish oily residue (3.48 g), which was submitted to column chromatography through silica gel (175 g, hexane / AcOEt, gradient elution). On elution with hexane / AcOEt 80:20, starting keto ester 7 (460 mg) was recovered. On elution with hexane / AcOEt 70:30, alcohol 9 (460 mg), a mixture of alcohols 9 / 8 (1.03 g) in a ratio 2:3 (¹H NMR), and alcohol 8 (1.14 g) were successively separated (50% total yield of 8 and 25% total yield of 9; 58% and 29%, respectively, based on recovered keto ester 7).

3a,4,6,7-Tetrahydro-9-methyl-3a,7-[1]propeno-2H-indazole-3,5-dione (11). A mixture of keto ester 7 (400 mg, 1.50 mmol), KOH pellets (5.19 g, 92.6 mmol), 80% aqueous hydrazine monohydrate (5.30 mL, 85.0 mmol) and diethylene glycol (20 mL) was heated at 120 °C for 2 h. diluted with diethylene glycol (10 mL) and heated at 140 °C for 15 h. The mixture was allowed to cool to room temperature, poured into a water-crushed ice mixture (75 g), made acidic to pH 1 with conc. HCl (4 mL) and extracted with CH₂Cl₂ (3 × 50 mL). The combined organic extracts were washed with brine (50 mL), dried with anhydrous Na₂SO₄ and concentrated in vacuo to give a white solid residue (270 mg), which was taken up in dioxane (4 mL) and treated with 2 N HCl for 3.5 h. The organic solvent was evaporated at reduced pressure and the aqueous layer was extracted with CH₂Cl₂ (3 × 3 mL). The combined organic extracts were washed with saturated aqueous NaHCO3 (3 × 4 mL), dried with anhydrous Na₂SO₄ and concentrated in vacuo, affording essentially pure 11 (124 mg, 40% yield) as a white solid. H NMR (200 MHz, DMSO- d_6) δ 1.62 (s, 3 H, 9-CH₃), 2.40-2.50 (complex signal, 4 H) and 2.75 (d, J = 15.5 Hz, 2 H) (methylenic protons), 3.37 (s, NH), 3.39 (br. s, 1 H, 7-H), 5.59 (br. d, J =5.5 Hz, 1 H, 8-H). ¹³C NMR (50.3 MHz, DMSO-d₆) δ 22.0 (CH₃, 9-CH₃), 33.9 (CH, C7), 40.7 (CH₂, C10), 46.8 (CH₂, C6), 48.1 (CH₂, C4), 125.1 (CH, C8), 133.2 (C, C9), 164.4 (C, C7a), 179.3 (C, C3), 206.1 (C, C5). The signal corresponding to C3a was not observed.

(7,7-Ethylenedioxy-anti-9-hydroxy-3-methylbicyclo[3.3.1]non-3-en-1-yl)methanol (12). A mixture of keto ester 7 (400 mg, 1.50 mmol) and NaBH₄ (125 mg, 3.29 mmol) in MeOH (8 mL) was stirred at room temperature for 24 h and quenched with saturated aqueous NH₄Cl (40) mL). The organic solvent was evaporated at reduced pressure and the remaining aqueous phase was diluted with water (80 mL), made alkaline with 25% aqueous NH₄OH (30 mL), and extracted with CH₂Cl₂ (4 × 50 mL). The combined organic extracts were dried with anhydrous Na₂SO₄ and concentrated in vacuo to give a yellow oil (360 mg), which was submitted to column chromatography through silica gel (26 g, hexane / AcOEt, gradient elution). On elution with hexane / AcOEt 40:60, diol 12 (110 mg, 31% yield) was isolated. The analytical sample of 12 was obtained by sublimation at 90 °C / 1 Torr: white crystals, m.p. 102.5-105.5 °C, IR (KBr) v 3352, 2986, 2939, 2872, 2662, 1465, 1428, 1408, 1375, 1351, 1311, 1289, 1256, 1213, 1149. 1094, 1058, 1031, 998, 974, 947, 895, 878, 823, 786, 739, 666 cm⁻¹. MS, m/z (%): 240 (M·+, 9), 209 (M+ - CH₂OH, 14), 191 (M+ - CH₂OH - H₂O, 12), 178 (8), 161 (10), 160 (48), 153 (7), 152 (12), 149 (6), 148 (6), 147 (25), 121 (36), 120 (55), 119 (37), 109 (35), 107 (29), 105 (46). 95 (31), 93 (42), 91 (82), 87 (93), 86 (42), 81 (36), 80 (52), 79 (69), 77 (70), 73 (25), 69 (30), 67 (49), 65 (42), 59 (67), 57 (44), 55 (100), 53 (69), 51 (28). Anal. Calcd. for C₁₃H₂₀O₄: C, 64.97; H, 8.39. Found: C, 64.89; H, 8.56.

7,7-ethylenedioxy-anti-9-[(1-imidazolyl)thiocarbonyloxy]-3-Methyl methylbicyclo[3.3.1]non-3-ene-1-carboxylate (13). A solution of alcohol 8 (5.43 g, 20.3 mmol) and 1,1'-thiocarbonyldiimidazole (5.97 g, 33.5 mmol) in 1,2-dichloroethane (90 mL) was heated under reflux for 3 h. The cold mixture was partitioned between CH₂Cl₂ (200 mL) and water (200 mL). The organic layer was dried with anhydrous Na₂SO₄ and evaporated at reduced pressure, to give a brown oil (11.6 g), which was submitted to column chromatography through silica gel (40-60 µm, 325 g, hexane / AcOEt, gradient elution). On elution with hexane / AcOEt 40:60, compound 13 (5.42 g, 71% yield) was isolated. The analytical sample of 13 was obtained by recrystallization from acetonitrile: pale yellow crystals, m.p. 135-136 °C. IR (KBr) v 3446, 3171, 3122, 2958, 2937, 2883, 1733, 1683, 1654, 1642, 1531, 1515, 1465, 1439, 1388, 1376, 1356, 1324, 1280, 1263, 1245, 1226, 1160, 1141, 1092, 1055, 1025, 1007, 995, 984, 949. 940, 919, 890, 843, 831, 821, 807, 758, 712, 655 cm⁻¹. MS, m/z (%): 379 (7), 378 (M·+, 30), 345 (13), 311 (M+ - C₃H₃N₂, 19), 251 (M+ - C₄H₃N₂OS, 19), 250 (14), 219 (M+ - C₄H₄N₂OS - CH₃O, 14), 207 (13), 192 (6), 191 (M+ - C₄H₄N₂OS - COOMe, 42), 190 (5), 189 (5), 176 (7), 175 (40), 174 (7), 165 (15), 164 (21), 163 (8), 151 (6), 147 (M+ - C₄H₄N₂OS - COOMe -C₂H₄O, 67), 119 (44), 105 (100), 91 (74), 87 (70), 86 (44), 79 (29), 77 (31), 59 (44). Anal. Calcd. for C₁₈H₂₂N₂O₅S: C, 57.13; H, 5.86; N, 7.40; S, 8.47. Found: C, 57.17; H, 5.94; N, 7.55; S. 8.35.

Methyl 7,7-ethylenedioxy-3-methylbicyclo[3.3.1]non-3-ene-1-carboxylate (10).

From compound 13. A solution of Bu₃SnH (1.50 mL, 1.62 g, 5.57 mmol) in anhydrous toluene (100 mL) was treated dropwise over 30 min with a solution of compound 13 (1.00 g. 2.65 mmol) and AIBN (45.2 mg, 0.27 mmol) in anhydrous toluene (100 mL). The reaction mixture was heated under reflux for 1.5 h, cooled to room temperature and concentrated in vacuo, to give a yellow oily residue (2.76 g) which was taken up in acetonitrile (250 mL) and washed with hexane (2×100 mL). The acetonitrile solution was evaporated at reduced pressure. affording a colorless oil (0.92 g), which was submitted to column chromatography through silica gel (100 g, hexane / AcOEt, gradient elution). On elution with hexane / AcOEt 85:15, slightly impure 10 (320 mg) was isolated. This product was taken up in acetonitrile (100 mL) and washed with hexane (50 mL). Evaporation of the acetonitrile solution at reduced pressure afforded pure 10 (290 mg, 42% yield) as a colorless oil. The analytical sample of 10 was obtained as a colorless oil, by distillation at 80 °C / 0.5 Torr. IR (NaCl) v 2950, 2919, 1732. 1456, 1432, 1372, 1353, 1290, 1254, 1228, 1162, 1145, 1097, 1056, 1006, 985, 950, 869, 828 cm⁻¹. MS, m/z (%): 252 (M⁺+, 21), 237 (5), 221 (5), 220 (M⁺ - CH₃OH, 26), 194 (6), 193 (M⁺ -COOMe, 38), 192 (6), 191 (6), 177 (12), 176 (5), 175 (6), 170 (14), 165 (11), 164 (8), 152 (5), 151 (M+ - C₅H₉O₂, 29), 119 (36), 107 (54), 105 (45), 91 (91), 87 (36), 86 (30), 85 (58), 83 (89). 79 (46), 77 (40), 65 (27), 59 (100), 58 (29), 57 (43), 55 (48). Anal. Calcd. for C₁₄H₂₀O₄: C. 66.64; H, 8.00. Found: C, 66.61; H, 8.27.

From alcohol 9. Compound 14 was prepared in a similar manner to that described for compound 13, starting from a solution of alcohol 9 (2.00 g, 7.46 mmol) and 1.1'-thiocarbonyldiimidazole (2.19 g, 12.3 mmol) in 1,2-dichloroethane (30 mL) but the reaction was carried out by heating at 120 °C in a pressure flask for 3 h. The resulting brown oily residue (4.93 g) was submitted to column chromatography through silica gel (40-60 µm, 120 g, hexane /

AcOEt, gradient elution). On elution with hexane / AcOEt 50:50, starting alcohol 9 (1.00 g) was recovered. On elution with hexane / AcOEt 40:60, compound 14 (900 mg, 32% yield; 64% yield based on recovered 9) was isolated as a slightly impure material which could not be fully characterized owing to its instability and was used as such in the next step. Compound 10 was prepared from 14 as described from 13. From a solution of Bu₃SnH (0.30 mL, 324 mg, 1.11 mmol) in anhydrous toluene (20 mL) and a solution of compound 14 (200 mg, 0.53 mmol) and AIBN (9.0 mg, 0.055 mmol) in anhydrous toluene (20 mL), a colorless oil (200 mg) was obtained. This was submitted to column chromatography through silica gel (20 g, hexane / AcOEt, gradient elution). On elution with hexane / AcOEt 90:10, slightly impure 10 (110 mg) was isolated. This product was taken up in acetonitrile (50 mL) and washed with hexane (25 mL). Evaporation of the acetonitrile solution at reduced pressure afforded pure 10 (80 mg, 60% yield) as a colorless oil.

3-Methyl-7-oxobicyclo[3.3.1]non-3-ene-1-carboxylic acid (15). A mixture of ester 10 (1.18 g, 4.68 mmol), 20% aqueous NaOH (95 mL, 0.47 mol), THF (95 mL) and MeOH (95 mL) was heated under reflux for 48 h. The organic solvent was evaporated *in vacuo* and the remaining aqueous phase was diluted with water (50 mL), washed with CH_2Cl_2 (2 × 75 mL), made acidic to pH 1 with 5 N HCl (90 mL), and extracted with CH_2Cl_2 (5 × 75 mL). The combined organic extracts were washed with brine (100 mL), dried with anhydrous Na₂SO₄ and evaporated at reduced pressure, to give a yellow oil (720 mg) which solidified on standing. Recrystallization of this solid from AcOEt (5 mL) afforded pure keto acid 15 (490 mg, 54% yield): white crystals, m.p. 153-155 °C. IR (KBr) v 3600-2350 (max. at 3423, 3048, 2955, 2928, 2853, 2643, 2501, 2363), 1701, 1449, 1427, 1405, 1303, 1282, 1261, 1219, 1182, 1079, 1061, 1038, 973, 922, 887, 864, 844, 828, 722, 662 cm⁻¹. MS, m/z (%): 194 (M·+, 18), 176 (M·+-H₂O, 19), 149 (M+ - COOH, 6), 137 (M+ - C₃H₅O, 45), 93 (100), 91 (80), 79 (34), 77 (50). Anal. Calcd. for $C_{11}H_{14}O_3$: C, 68.02; H, 7.27. Found: C, 67.98; H, 7.34.

Methyl *N*-{3-methyl-7-oxobicyclo[3.3.1]non-3-en-1-yl}carbamate (16). A solution of keto acid 15 (1.63 g, 8.40 mmol), Et₃N (1.17 mL, 0.85 g, 8.39 mmol) and diphenylphosphoryl azide (1.71 mL, 2.18 g, 7.91 mmol) in anhydrous chlorobenzene (29 mL) was heated at 90 °C for 3.5 h. Anhydrous MeOH (46 mL) was added to the cold solution and the resulting mixture was heated under reflux for 17 h. The solvent was evaporated at reduced pressure to give a brown oil (5.60 g), which was submitted to column chromatography through silica gel (157 g, CH₂Cl₂ / AcOEt, gradient elution). On elution with CH₂Cl₂ / AcOEt 85:15, keto carbamate 16 (1.74 g, 99% yield based on diphenylphosphoryl azide) was isolated as a light yellow oil which solidified on standing. The analytical sample of 16 was obtained by sublimation at 110 °C / 1 Torr: white solid, m.p. 65.5-66.5 °C. IR (KBr) v 3324, 3054, 3038, 2985, 2961, 2941, 2881, 2841, 1727. 1709, 1674, 1536, 1463, 1436, 1415, 1363, 1342, 1296, 1250, 1222, 1193, 1133, 1112, 1073. 1041, 1032, 927, 816, 779, 716, 602 cm⁻¹. MS, m/z (%): 223 (M·+, 8), 167 (10), 166 (M+-C₃H₅O, 93), 165 (10), 134 (M+-C₃H₆O-MeO, 100), 107 (26), 106 (M+-C₃H₆O-COOMe, 59), 91 (84), 79 (42), 77 (35), 59 (50), 53 (26). Anal. Calcd. for C₁₂H₁₇NO₃: C, 64.55; H, 7.68; N, 6.27. Found: C, 64.50; H, 7.75; N, 6.30.

5,6,9,10-Tetrahydro-5-(methoxycarbonylamino)-7-methyl-5,9-methano-1*H*-cycloocta[*b*]pyridin-2-one (19) and 5,8,9,10-tetrahydro-9-(methoxycarbonylamino)-7-

methyl-5,9-methano-1H-cycloocta[b]pyridin-2-one (20). A mixture of keto carbamate 16 (950 mg, 4.26 mmol), pyrrolidine (0.44 mL, 0.38 g, 5.32 mmol), 4 Å molecular sieves (2.5 g) and anhydrous benzene (50 mL) was heated under reflux for 5 h. The mixture was cooled to room temperature and was filtered under argon. The filtrate was treated with propiolamide (0.88 g. 12.8 mmol) and the reaction mixture was heated under reflux for 15 h. The resulting red suspension was allowed to cool to room temperature, diluted with MeOH (200 mL) and filtered. The filtrate was evaporated at reduced pressure to give a red solid residue (2.58 g), which was taken up in AcOEt (600 mL) and extracted with 1 N NaOH (5 \times 150 mL). The combined aqueous extracts were neutralized with 0.75 N HCl (ca. 1 L) and extracted successively with CH_2Cl_2 (5 × 100 mL) and AcOEt (5 × 100 mL). The combined organic extracts were washed with saturated aqueous NaHCO₃ (2×50 mL), dried with anhydrous Na₂SO₄ and evaporated at reduced pressure, to give a yellow solid residue (450 mg), which was submitted to column chromatography through silica gel (40-60 µm, 45 g, hexane / AcOEt / MeOH, gradient elution). On elution with AcOEt / MeOH 95:5, a mixture of pyridones 19 and 20 (240 mg) in a ratio 9:16 (1H NMR) was separated. On elution with AcOEt / MeOH 90:10, a mixture of pyridones 19 and 20 (110 mg) in a ratio 2:1 (¹H NMR) and pure 19 (40 mg) were isolated (33% total yield of pyridones; 17% yield of 19 and 16% yield of 20). Meanwhile, the initial AcOEt solution was dried with anhydrous Na₂SO₄ and evaporated at reduced pressure, to give a residue (1.54 g) which was submitted to column chromatography through silica gel (45 g, CH₂Cl₂ / AcOEt. gradient elution). On elution with CH₂Cl₂ / AcOEt 85:15, starting keto carbamate 16 (380 mg) was recovered (55% total yield of pyridones based on recovered 16; 28% yield of 19 and 27% yield of 20).

Medium pressure liquid chromatography (MPLC) separation of a mixture of 19 and 20. The chromatographic separation of pyridones 19 and 20 was carried out by MPLC. The mixture of pyridones (300 mg of a mixture of 19 and 20 in the ratio of 8:7) was introduced in one portion dissolved in MeOH (8 mL). Mixtures of AcOEt / MeOH (gradient elution) were used as eluent, with a flow of 14 mL / min and an initial pressure of 4 bar. On elution with AcOEt / MeOH 95:5, pure pyridone 20 (110 mg) and a mixture of 19 and 20 (60 mg) in a ratio 2:1 were successively isolated. On elution with AcOEt / MeOH 94:6, pure pyridone 19 (110 mg) was isolated. The analytical samples of 19 and 20 were obtained by recrystallization from AcOEt / MeOH 9:2 and AcOEt / MeOH 5:2, respectively.

Spectroscopic and analytical data of pyridone *19*: white crystals, m.p. 266-269 °C (dec.). IR (KBr) v 3446, 3245, 3056, 2975, 2928, 2903, 2412, 2367, 1702, 1664, 1613, 1587, 1549, 1458, 1427, 1390, 1358, 1323, 1300, 1281, 1264, 1249, 1197, 1178, 1145, 1136, 1121, 1084, 1053, 1028, 925, 848, 819, 793, 781, 734, 685 cm⁻¹. ¹H NMR (CD₃OD) δ 1.59 (s, 3 H, 7-CH₃), 1.71 (ddd, J = 12.0 Hz, J' = 4.0 Hz, J'' = 1.5 Hz, 1 H, 11-H_{anti}), 1.96 (br. d, J = 16.5 Hz, 1 H, 6-H_{endo}), 2.41 (dm, $J \approx 16.5$ Hz, 1 H, 6-H_{exo}), overlaps in part 2.42 (ddd, J = 17.5 Hz, J' = J'' = 1.5 Hz, 1 H, 10-H_{endo}), 2.57 (br. d, $J \approx 12.0$ Hz, 1 H, 11-H_{syn}), 2.76 (m, 1 H, 9-H), 2.90 (dd, J = 17.5 Hz, J' = 5.0 Hz, 1 H, 10-H_{exo}), 3.56 (s, 3 H, NHCOOCH₃), 4.87 (s, NHCOOCH₃ + 1-H + H₂O), 5.51 (dm, J = 4.0 Hz, 1 H, 8-H), 6.35 (d, J = 9.5 Hz, 1 H, 3-H), 7.56 (d, J = 9.5 Hz, 1 H, 4-H). ¹³C NMR (CD₃OD + D₂O) δ 23.1 (CH₃, 7-CH₃), 31.3 (CH, C9), 34.1 (CH₂, C10), 34.4 (CH₂, C11), 45.1 (CH₂, C6), 52.2 (CH₃, NHCOOCH₃), 53.4 (C, C5), 117.9 (CH, C3), 122.4 (C, C4a), 125.7 (CH, C8), 133.4 (C, C7), 141.4 (CH, C4), 144.1 (C, C10a), 157.5 (C, NHCOOCH₃).

165.8 (C, C2). MS, m/z (%): 274 (M·+, 10), 259 (M+ - CH₃, 16), 232 (7), 215 (7), 206 (10), 205 (7), 200 (15), 199 (M·+ - NH₂COOMe, 20), 198 (11), 187 (8), 185 (5), 184 (M·+ - NHCOOMe - CH₃, 17), 174 (6), 173 (5), 170 (5), 166 (6), 160 (5), 159 (5), 158 (5), 109 (100). Anal. Calcd. for C₁₅H₁₈N₂O₃·1/3H₂O: C, 64.27; H, 6.72; N, 9.99. Found: C, 64.25; H, 6.52; N, 10.09.

Spectroscopic and analytical data of pyridone 20: white crystals, m.p. 260-262 °C (dec.). IR (KBr) v 3413, 3267, 3119, 3035, 2969, 2944, 2920, 2896, 2871, 2766, 2422, 2361, 2337, 1713, 1651, 1621, 1557, 1536, 1460, 1405, 1378, 1283, 1233, 1188, 1112, 1092, 1070, 1036, 887, 827, 782, 748, 653 cm⁻¹. ¹H NMR (CD₃OD) δ 1.60 (s, 3 H, 7-CH₃), 1.95 (dd, J = 11.5 Hz, J' = 3.0 Hz, 1 H) and 2.16 (br. d, J = 11.5 Hz, 1 H) (11-H_{syn} and 11-H_{anti}), 2.38 (br. d, J = 17.5 Hz. 1 H, 8-H_{exo}), 2.53 (br. d, J = 17.5 Hz, 1 H, 8-H_{endo}), 3.01 (br. d, J = 19.0 Hz, 1 H, 10-H_{endo}), 3.18 (br. d, J = 19.0 Hz, 1 H, 10-H_{exo}), 3.22 (m, 1 H, 5-H), 3.61 (s, 3 H, NHCOOCH₃), 4.86 (s, NHCOOCH₃ + 1-H), 5.64 (d, J = 5.0 Hz, 1 H, 6-H), 6.27 (d, J = 9.0 Hz, 1 H, 3-H), 7.35 (d, J = 9.0 Hz, 1 H, 4-H). ¹³C NMR (CD₃OD + D₂O) δ 22.9 (CH₃, 7-CH₃), 35.2 (CH₂, C11), 35.3 (CH, C5), 41.1 (CH₂, C10), 46.3 (CH₂, C8), 51.4 (CH₃, NHCOOCH₃), 52.5 (C, C9), 116.7 (CH, C3), 123.2 (C, C4a), 126.6 (CH, C6), 132.4 (C, C7), 143.3 (C, C10a), 143.6 (CH, C4), 158.4 (C, NHCOOCH₃), 165.4 (C, C2). MS, m/z (%): 274 (M·+, 33), 207 (5), 200 (11), 199 (M·+ NH₂COOMe, 33), 198 (23), 187 (5), 184 (M·+ NHCOOMe - CH₃, 28), 160 (11), 159 (22), 109 (58), 59 (100), 55 (32). Anal. Calcd. for C₁₅H₁₈N₂O₃: C, 65.67; H, 6.62; N, 10.21. Found: C, 65.68; H, 6.69; N, 10.24.

5-Amino-5,6,9,10-tetrahydro-7-methyl-5,9-methano-1H-cycloocta[b]pyridin-2-one (5). A mixture of pyridone 19 (90 mg, 0.33 mmol), anhydrous HMPA (0.6 mL), and lithium 1propanethiolate (ca. 0.5 M solution in anhydrous HMPA [52], 4.40 mL, ca. 2.20 mmol) was heated at 40 °C for 24 h. The cold (room temperature) mixture was poured into ice (30 g) and concentrated in vacuo. The resulting brown residue (1.09 g) was submitted to column chromatography through silica gel (40-60 µm, 140 g, CHCl₃ / MeOH, gradient elution). On elution with CHCl₃ / MeOH 90:10, starting 19 (20 mg) was recovered. On elution with CHCl₃ / MeOH 60:40, pure amine 5 (45 mg, 63% yield; 81% yield based on recovered 19) was isolated. The analytical sample of 5 was obtained by recrystallization from AcOEt / MeOH 7:2: white crystals, m.p. 259-260 °C (dec.). IR (KBr) v 3442, 3392, 3307, 3261, 3173, 3076, 3031, 2951. 2939, 2887, 2818, 2722, 2647, 1644, 1606, 1554, 1461, 1426, 1413, 1295, 1263, 1191, 1177. 1125, 1114, 1070, 1036, 1017, 1000, 987, 968, 953, 888, 876, 850, 686 cm⁻¹. ¹H NMR $(CD_3OD) \delta 1.58 \text{ (s, 3 H, 7-CH_3)}, 1.78 \text{ (dm, } J = 12.0 \text{ Hz, 1 H, 11-H}_{syn}), 1.84 \text{ (ddd, } J = 12.0 \text{ Hz, } 1.00 \text{ Hz}$ J' = 4.5 Hz, J'' = 1.5 Hz, 1 H, 11-H_{anti}), 1.96 (br. d, J = 17.0 Hz, 1 H, 6-H_{endo}), 2.27 (dm, J = 1.5 Hz) 17.0 Hz, 1 H, 6-H_{exo}), 2.44 (dm, J = 17.0 Hz, 1 H, 10-H_{endo}), 2.75 (m, 1 H, 9-H), 2.87 (dd, J = 17.0 Hz, 1 H, 10-H_{endo}) 17.0 Hz, J' = 5.5 Hz, 1 H, 10-H_{exo}), 4.88 (s, NH₂ + 1-H), 5.47 (dm, J = 4.5 Hz, 1 H, 8-H), 6.38 (d, J = 9.0 Hz, 1 H, 3-H), 7.84 (d, J = 9.0 Hz, 1 H, 4-H). ¹³C NMR (CD₃OD) δ 23.1 (CH₃, 7-CH₃), 31.5 (CH, C9), 34.6 (CH₂, C10), 39.1 (CH₂, C11), 47.3 (CH₂, C6), 50.3 (C, C5), 117.9 (CH, C3), 124.1 (C, C4a), 125.2 (CH, C8), 134.8 (C, C7), 141.8 (CH, C4), 143.9 (C, C10a), 165.7 (C, C2). Exact mass calcd. for C₁₃H₁₆N₂O 216.1263, obsd. 216.1258.

9-Amino-5,6,9,10-tetrahydro-7-methyl-5,9-methano-1*H*-cycloocta[*b*]pyridin-2-one (21). To a suspension of carbamate 20 (96.0 mg, 0.35 mmol) in CHCl₃ (13 mL), trimethylsilyl iodide (0.50 mL, 0.70 g, 3.51 mL) was added dropwise and the mixture was heated under reflux for 8 h.

MeOH (13 mL) was then added and the reaction mixture was heated under reflux for 14 h. Evaporation of the solvents at reduced pressure gave a brown residue (270 mg) which was submitted to column chromatography through ammonia-saturated silica gel (22 g, CHCl₃ / MeOH, gradient elution). On elution with CHCl₃ / MeOH 96:4, amine 21 was obtained slightly contaminated with the non-isomerized amine (80 mg). This product was again submitted to column chromatography under the same conditions. On elution with CHCl₃ / MeOH 96:4, pure 21 (50 mg, 66% yield) was isolated. The analytical sample of 21 was obtained by recrystallization from AcOEt / MeOH 4:1: white crystals, m.p. > 300 °C (dec.). IR (KBr) v 3430, 2926, 1652, 1609, 1558, 1446, 1420, 1375, 1159, 1117, 835 cm⁻¹. ¹H NMR (CD₃OD) δ 1.63 (s, 3 H, 7-CH₃), 1.86-1.91 (complex signal, 2 H, 6-H_{endo} and 11-H_{anti}), 1.97 (dm, J = 12.0 Hz. 1 H, 11-H_{SVn}), 2.47 (ddm, J = 18.0 Hz, J' = 3.5 Hz, 1 H, 6-H_{exo}), 2.69 (d, J = 17.0 Hz, 1 H, 10- H_{endo}), 2.80 (d, J = 17.0 Hz, 1 H, 10-H_{exo}), 3.17 (m, 1 H, 5-H), 4.85 (s, NH₂ + 1-H), 5.35 (br. s. 1 H, 8-H), 6.38 (d, J = 9.0 Hz, 1 H, 3-H), 7.45 (d, J = 9.0 Hz, 1 H, 4-H). ¹³C NMR (CD₃OD) δ 23.2 (CH₃, 7-CH₃), 33.7 (CH, C5), 36.5 (CH₂, C11), 38.3 (CH₂, C6), 40.4 (CH₂, C10), 51.2 (C. C9), 118.5 (CH, C3), 120.1 (C, C4a), 126.3 (CH, C8), 137.0 (C, C7), 142.1 (C, C10a), 144.8 (CH, C4), 165.7 (C, C2). Exact mass calcd. for C₁₃H₁₆N₂O 216.1263, obsd. 216.1259.

X-ray Crystal-Structure Determination of 9 [53]. A prismatic crystal was selected and mounted on a Enraf-Nonius CAD4 four-circle diffractometer. Unit-cell parameters were determined by automatic centering of 25 reflections ($12 < \theta < 21^{\circ}$) and refined by the least-squares method. Intensities were collected with graphite-monochromatized Mo- $K\alpha$ radiation, using $\omega/2\theta$ scan technique. 1720 reflections were measured in the range $2.23 \le \theta \le 29.96$, 1647 of which were non-equivalent by symmetry [Rint (on I) = 0.070]. 976 reflections were assumed

Table 3
Experimental data of the X-ray crystal structure determination of 9.

| Molecular formula | $C_{14}H_{20}O_{5}$ | F(000) | 572 |
|---------------------|---------------------|---|-----------------------------|
| Molecular mass | 268.32 | d (calcd) [Mg m ⁻³] | 1.324 |
| Temperature | 293(2)K | Size of crystal [mm] | $0.1 \times 0.1 \times 0.2$ |
| Crystal system | Monoclinic | Measured reflections | 1720 |
| Space grup | P2 ₁ /n | Independent reflections | 1647 |
| Cell parameters | [a] | Observed reflections | 976 |
| a [Å] | 11.57(3) | $\mu(Mo-K\alpha)$ [mm ⁻¹][b] | 0.100 |
| b [Å] | 8.747(12) | R | 0.076 |
| c [Å] | 13.32(2) | Rw | 0.180 |
| α [°] | 90 | $\Delta \rho_{\text{max}}^{\text{[c]}}$ (eÅ ⁻³) | 0.228 |
| β[°] | 95.6(2) | $\Delta \rho_{\min}^{[d]}$ (eÅ-3) | -0.209 |
| γ[°] | 90 | Refined parameters | 176 |
| V [Å ³] | 1341(4) | Max. shift / e.s.d. | 0.001 |
| Z | 4 | | |

[[]a] Determined by automatic centering of 25 reflections ($12 \le \theta \le 21^{\circ}$).

[[]b] $\mu(\text{Mo-}K\alpha)$, Linear absorption coefficient. Radiation Mo- $K\alpha$ ($\lambda = 0.71069\text{Å}$).

[[]c] Maximum and [d] minimum peaks in final difference synthesis.

as observed by applying the condition $I > 2\sigma(I)$. Three reflections were measured every two hours as orientation and intensity control; significant intensity decay was not observed. Lorentz polarization but no absorption corrections were made. The structure was solved by Direct methods, using the SHELXS computer program [54] and refined by the full-matrix least-squares method with the SHELX-93 computer program [55] using 1597 reflections (very negative intensities were not assumed). The function minimized was Σ w ($|F_0|^2 - |F_c|^2$), where $W = [\sigma^2(I) + (0.0814P)^2]^{-1}$, and $P = (|F_0|^2 + 2|F_c|^2) / 3$. f, f and f" were taken from International Tables of X-ray Crystallography [56]. All H atoms were computed and refined with an overall isotropic temperature factor using a riding model. Goodness of fit on $F^2 = 1.128$ for all observed reflections. Mean shift / e.s.d. = 0.001.

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